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J. Nat. Prod., 1992, 55 (7), 967-969• DOI: 10.1021/np50085a021 • Publication Date (Web): 01 July 2004

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Chemical Society. 1155 Sixteenth Street N.W., Washington, DC 20036

NAKAHALENE AND CYTOTOXIC PRINCIPLES OF FORMOSAN *RHAMNUS* SPECIES¹

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ABSTRACT.—A new naphthalene compound, named nakahalene, has been isolated from the root bark of *Rhamnus nakaharai* (Rhamnaceae) together with chrysophanol, physcion, emodin, rhamnocitrin, and kaempferol. Nakahalene has been characterized as 2-acetyl-3-methyl-6methoxylnaphthalene-1,8-diol [4]. The cytotoxic effects of the isolated anthraquinones of *Rhamnus formosana* and *Rhamnus nakaharai* were investigated.

In a previous paper (1), we reported the isolation of a new flavonol triglycoside, rhamnustrioside, and kaempferol from the fresh fruits of *Rhamnus nakaharai* Hayata (Rhamnaceae). In continuation of this work, the known compounds chrysophanol [1], physcion [2], emodin [3], rhamnocitrin, and kaempferol, and a new naphthalene compound, nakahalene [4], were isolated from the fresh root bark of this same plant. In this paper, we report the isolation and characterization of this new naphthalene compound 4. Because some of the an-

TABLE 1.	Cytotoxicity	of Anthrac	quinones	Against	Various	Tumor	Cell Lines.

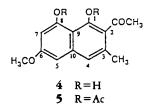
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R ₁ OR ₂ O OH Me	1 $R_1 = R_2 = H$ 2 $R_1 = OMe, R_2 = H$ 3 $R_1 = OH, R_2 = H$ 6 $R_1 = ORha, R_2 = H$ 7 $R_1 = OMe, R_2 = GlcORha$			
Compounds	$ED_{50}(\mu g/ml)(N-8)$			
	PLC/PRF/5 KB			
1	3.23 5.68 1.44 1.55 5.91 3.46 0.80 2.49 2.50 3.58 5.29 0.16			

^aFor significant activity of the pure compound, an ED₅₀ \leq 4.0 µg/ml is required (10).

thraquinones exhibited cytotoxic effects against HL-60 cells (2), compounds 1-3, frangulin B [6], and physcion 8-0rhamnosyl-($1\rightarrow 2$)-glucoside [7] were

¹Part VI in the series "The Constituents of Formosan *Rhamnus* Species." For part V see C.N. Lin, M.I. Chung, K.H. Gan, and C.M. Lu, *Phytochemistry.* **30**, 3101 (1991).



screened for cytotoxic effects against human hepatoma PLC/PRF/5 and KB cell lines. Compounds **6** and **7** were isolated from *Rhamnus formosana* Matsum (3).

RESULTS AND DISCUSSION

Compound 4 showed blue fluorescence under uv. Its uv and ir spectra suggested the presence of a hydroxynaphthalene skeleton (4). Its ¹H-nmr spectrum indicated the presence of an aromatic Me signal at δ 2.61, an acetyl signal at δ 2.72, an MeO signal at δ 3.87, a pair of 1H doublet signals (meta coupling) at δ 6.47 (d, J = 2.4 Hz, H-7), and 6.48 (d, J = 2.4 Hz, H-5), a 1H singlet signal at δ 6.83 (H-4) and two chelated phenolic hydroxyl signals at δ 10.42 (1H, s, exchangeable with D₂O, 8-OH) and 17.75 (1H, s, exchangeable with D_2O_1 , 1-OH) (4,5). The ¹H nmr spectrum of 4 diacetate [5] showed the presence of two singlets for acetyl signals at δ 2.31 and 2.36, an aromatic Me signal at δ 2.38, an acetyl signal at δ 2.50, an MeO signal at δ 3.89, a pair of 1H doublet signals (meta coupling) at δ 6.80 (d, J = 2.4 Hz, H-7) and 6.98 (d, J = 2.4Hz, H-5) and a 1H singlet signal at δ 7.47. The downfield shift of H-4, H-5, and H-7 of **4** by acetylation supported the characterization of 4.

The assignments of ¹³C-nmr spectra of 4 were obtained by ¹H-decoupling spectra and DEPT pulse sequence and assigned by comparison with data reported in the literature (6,7). The presence of peaks at m/z 127 and 128 in the eims of 4 also clearly indicated that 4 possessed the naphthalene skeleton (6). Based on the above evidence, compound **4** was characterized as 2-acetyl-3-methyl-6-methoxynaphthalene-1,8-diol.

The cytotoxic effects of 1-3, 6, and 7 against human hepatoma PLC/PRF/5 and KB cells were studied (8,9), and the results are listed in Table 1. Compound 6 exhibited the most significant cytotoxic effects against these cell lines. From Table 1, it is clearly indicated that 0-glycosylation or methylation of the C₆-OH of 3 enhances the cytotoxic effect against human hepatoma PLC/PRF/5 cells in vitro.

EXPERIMENTAL

GENERAL EXPERIMENTAL PROCEDURES.— All mp's are uncorrected. Ft nmr spectra were recorded on a VXR-300/51 Superconducting High Resolution FT NMR system; ir spectra on a Hitachi model 260-30; and ms on a JMS-HX 110 Mass Spectrometer.

EXTRACTION AND SEPARATION.—The fresh root bark (0.7 kg) of *R. nakaharai* was collected at Ali, Wu-Tai Shian, Ping-Tung Hsien, Taiwan, during July 1990, chipped, and extracted with hot MeOH. The MeOH extract was chromatographed on Si gel. Elution with cyclohexane- C_6H_6 (1:4) yielded **1** and **2**. Elution with C_6H_6 -EtOAc (5:1) yielded **3**. Elution with C_6H_6 cyclohexane-EtOAc (10:4:1) yielded **4**, kaempferol, and rhamnocitrin. Compounds **1–3**, kaempferol, and rhamnocitrin were identified by uv, ir, nmr, ms, and comparison of the mmp's and spectral data with those of the authentic samples.

2-ACETYL-3-METHYL-6-METHOXYNAPHTHA-LENE (NAKAHALENE) [4].—Yellow needles (EtOAc): mp 207–208°; ir ν max KBr cm⁻¹ 3350, 1640, 1580; uv λ max (MeOH) nm (log ϵ) 275 (3.95), 316 (3.17), 332 (sh) (3.02), 400 (3.36); ¹H nmr (CDCl₃) see text; ¹³C nmr (CDCl₃) δ 163.6 (C-1), 108.3 (C-2), 139.9 (C-3), 121.0 (C-4), 100.8 (C-5), 160.2 (C-6), 99.2 (C-7), 169.7 (C-8), 112.3 (C-9), 134.2 (C-10), 25.3 (Me), 31.7 (Ac), 55.4 (OMe), 203.5 (Ac); eims m/z (rel. int.) [M]⁺ 246 (62), 231 (100), 185 (12), 128 (10), 127 (9), 115 (12). Anal. calcd for C₁₄H₁₄O₄ 246.0891, found 246.0907.

COMPOUND **4** DIACETATE [**5**].—Colorless needles (MeOH): mp 190–191°; ir ν max KBr cm⁻¹ 1760, 1690, 1635; ¹H nmr (CDCl₃) see text; ¹³C nmr (CDCl₃) δ 142.9 (C-1), 115.9 (C-2), 138.5 (C-3), 127.8 (C-4), 114.7 (C-5), 159.7 (C-6), 105.6 (C-7), 147.8 (C-8), 133.4 (C-9), 133.8 (C-10), 19.8 (Me), 21.4, 21.6 (OAc), 32.5 (Ac), 55.8 (OMe), 170.5, 170.7 (OAc),

205.4 (Ac); eims m/z (rel. int.) [M]⁺ 330 (15), [M-42]⁺ 288 (18), [288-42]⁺ 246 (100), 231 (47), 115 (1). *Anal.* calcd for C₁₈H₁₈O₆ 330.1102, found 330.1114.

BIOLOGICAL ASSAYS.—PLC/PRF/5 cells were established from human hepatoma and are known to produce HBs Ag continuously in culture fluids (5). The cells were grown as continuous cultures in a growth medium consisting of Dulbecco's modified Eagle medium (DMEM, GIBCO, Grand Island, NY), 10% fetal bovine serum (FBS, GIBCO), 100 IU/ml penicillin, 100 μ g/ml streptomycin, and 2 mM L-glutamine. The KB cells were maintained on DMEM (GIBCO) containing with 10% FBS, L-glutamine and antibiotics. For microassays, the growth medium was supplemented further with 10 mM Hepes buffer, pH 7.3. The microassay for anticellular effect was performed as previously described (6,7).

ACKNOWLEDGMENTS

This work was supported partly by grants from the National Science Council of the Republic of China (NSC 78-0420-B037-17 and NSC 79-0420-B037-30) and National Research Institute of Chinese Medicine, Taipei, Taiwan, Republic of China.

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Received 24 September 1991